



Computational Biology LU 2014

Exercise 2 - Microarray Analysis

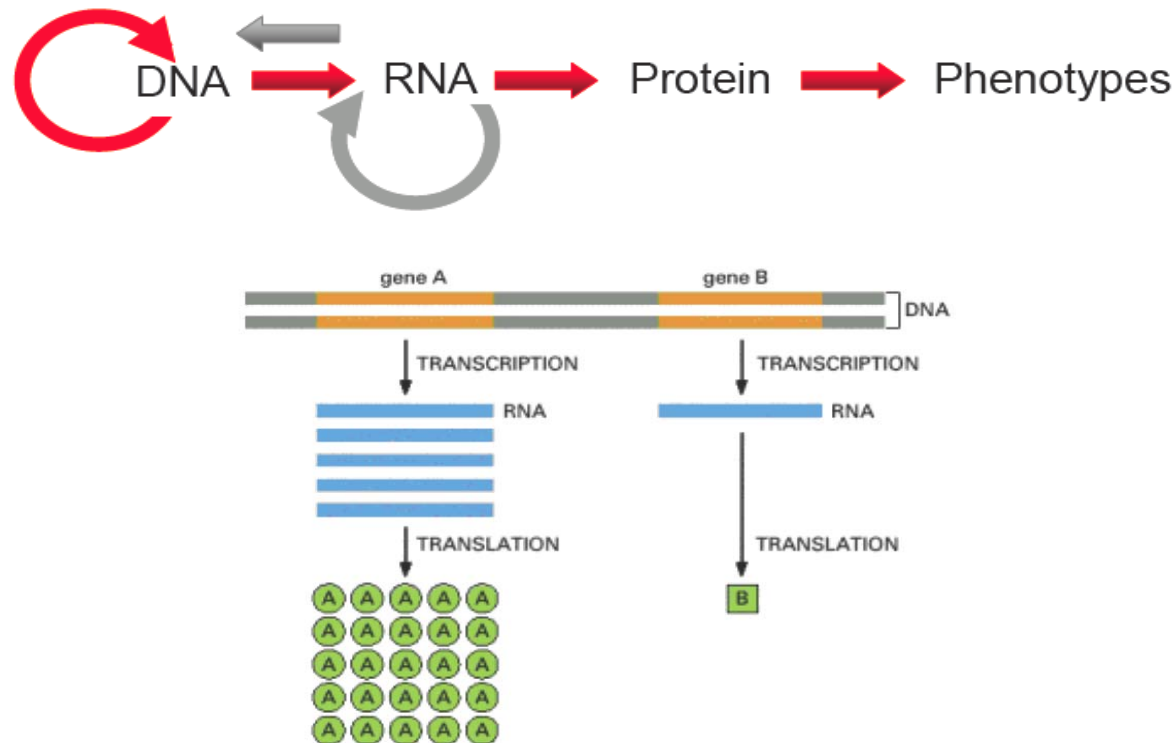
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Outline

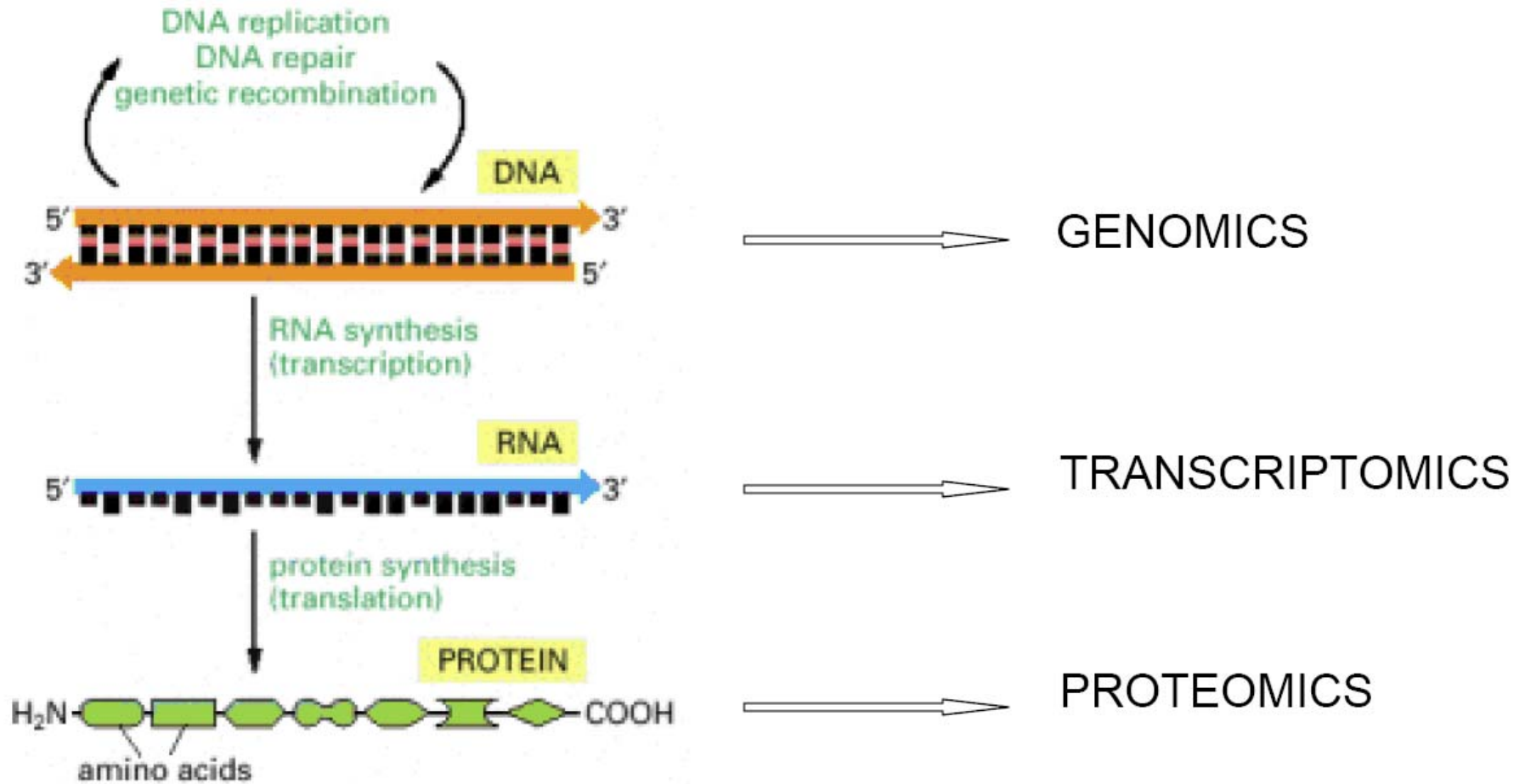
- Microarrays
 - Central Dogma of Molecular Biology
 - Omics
 - Two colour microarrays
 - Clustering
- Exercise 2 – help functions

Central Dogma of Molecular Biology





OMICS





Transcriptomics

- Transcriptomics = study of the transcriptome (parts of the genome that are transcribed)
- A gene is expressed when it is transcribed into RNA
- Genomes within and across species might be very similar
- The genes that are expressed is what makes the difference between individuals or between species



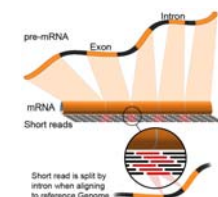
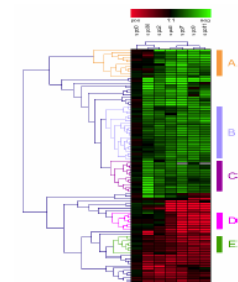
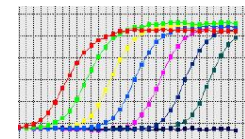
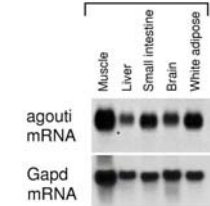
Transcriptomics

- The genome is static, the transcriptome is dynamic
- One gene has always the same sequence (except by mutations) but the same gene is differently expressed (at different rates) in different situations
- With transcriptomics we try to obtain a snap-shot of the cell transcriptional activity at a given time

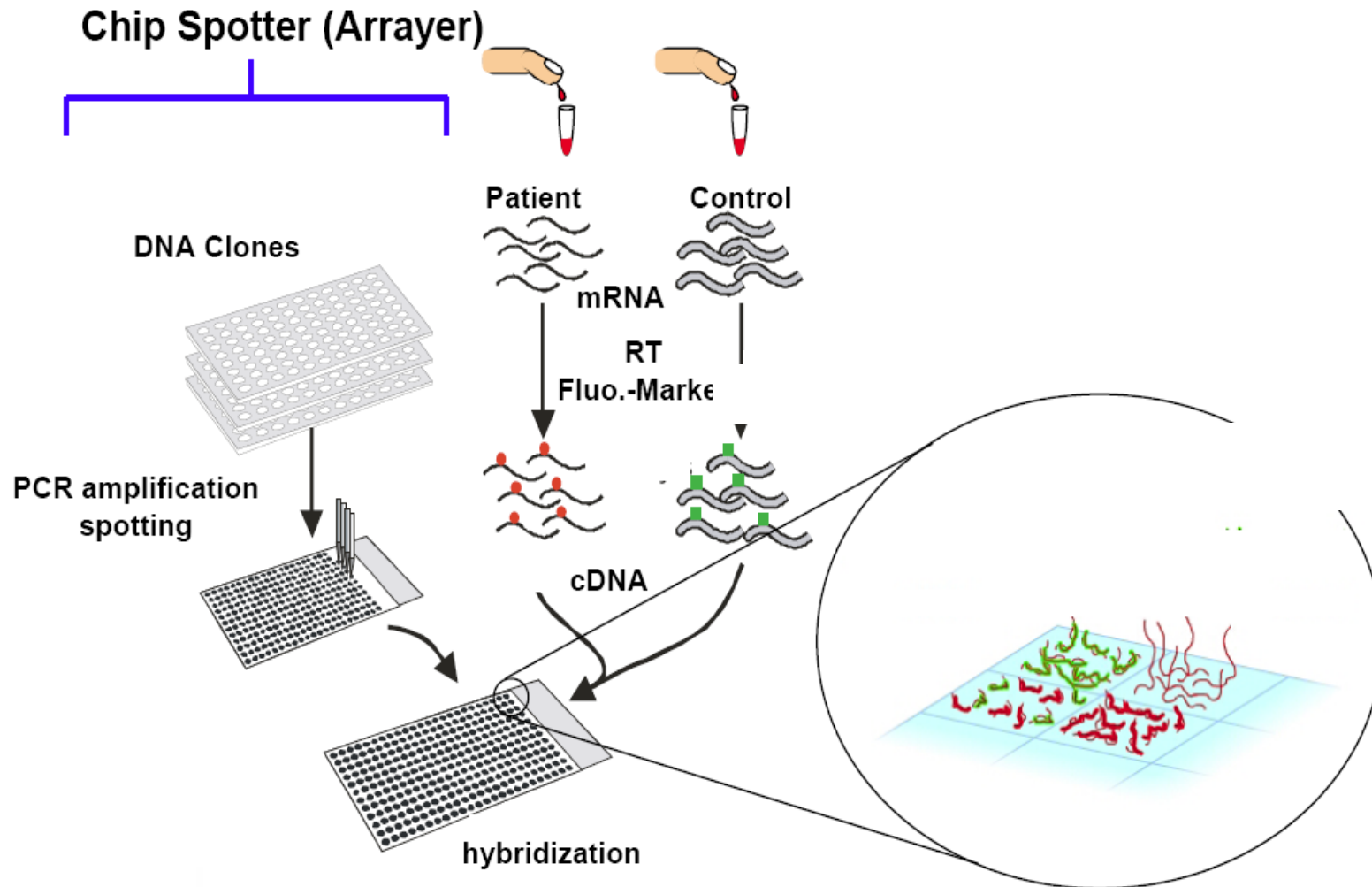


RNA expression profiling

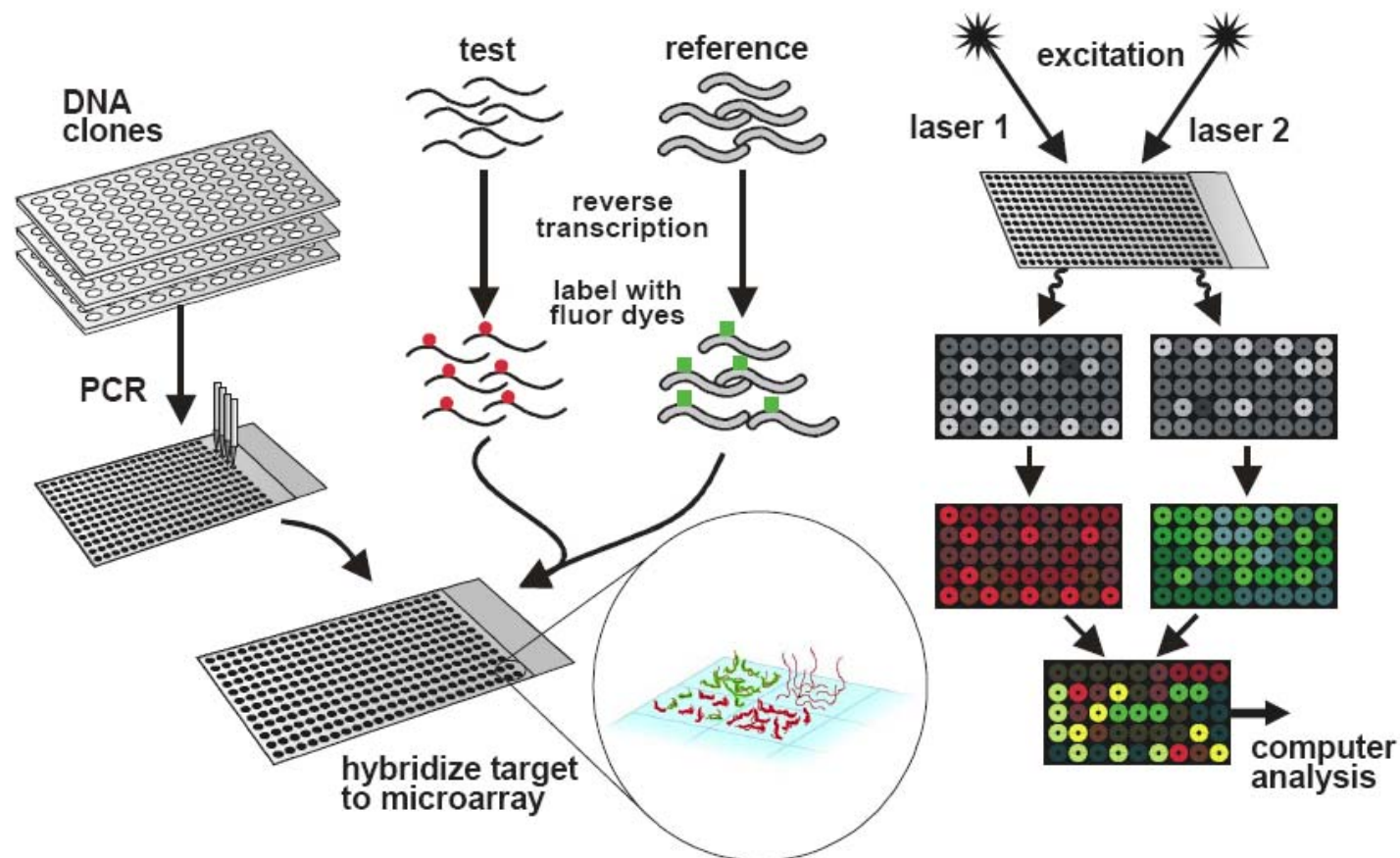
- Northern blotting
 - semi-quantitative
 - few genes
- Real time RT-PCR (qPCR)
 - medium throughput
- Microarray analysis
 - high throughput
 - 10.000-500.000 elements per chip
- RNA seq
 - high throughput
 - deep sequencing (short reads 25bp)



Two-color microarrays



Two-color microarrays





Analytical pipeline

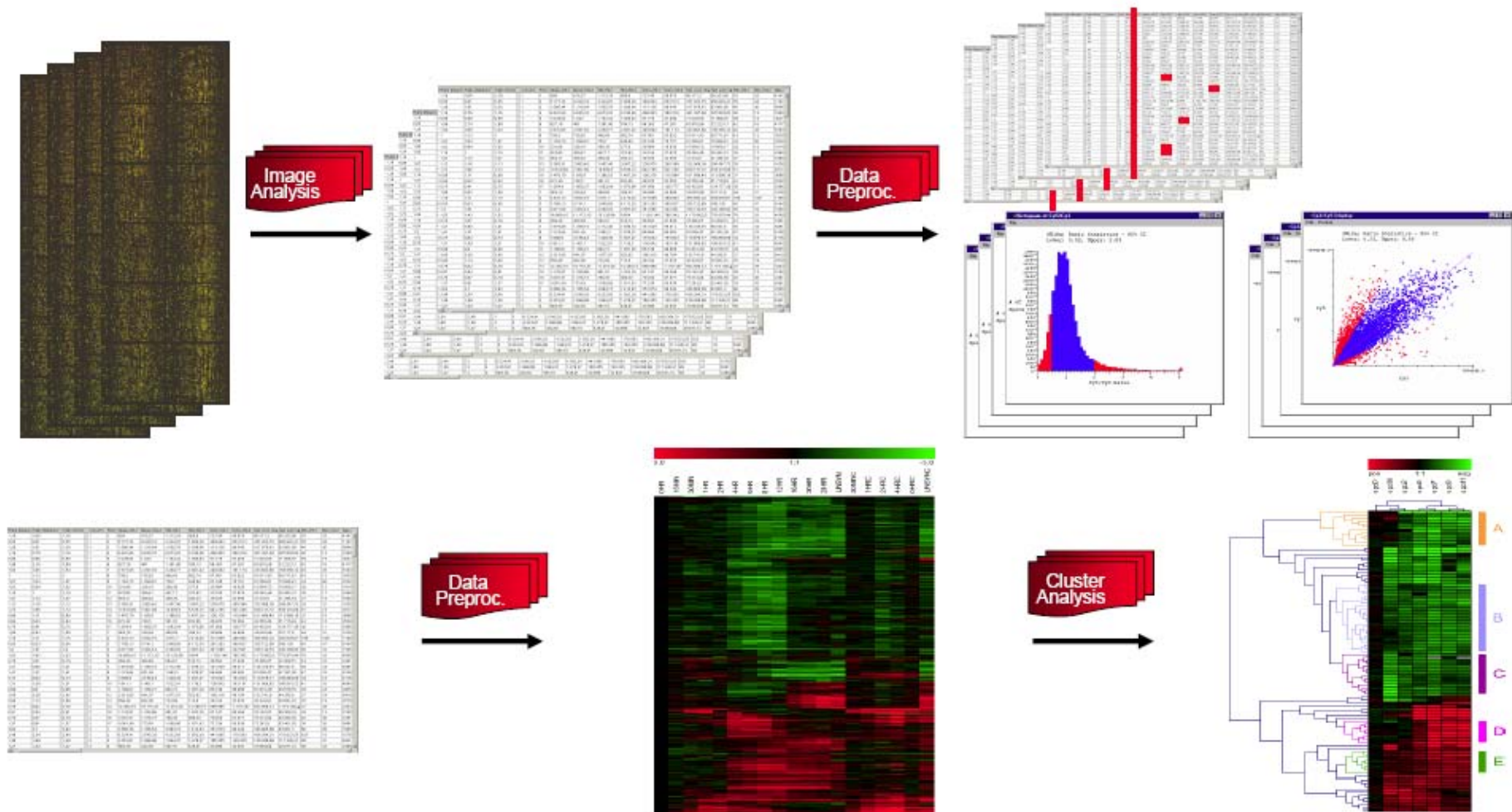
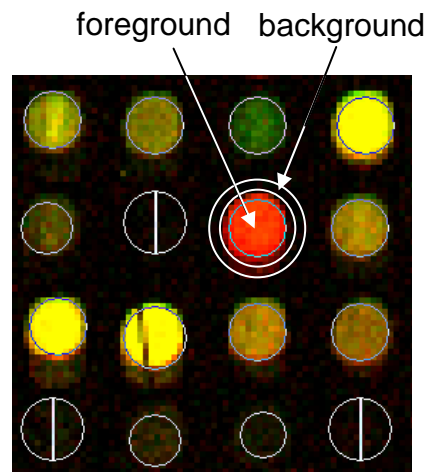


Image analysis and background correction

- Software available (GenePix, ImaGene, Agilent)
- Steps:
 - Gridding, assigns coordinates and gene information to the different spots
 - Segmentation: Foreground vs background
 - Intensity extraction



$$G_k = F532 \text{ mean} - B532$$

$$R_k = F635 \text{ mean} - B635$$

A lot of other parameter:
F635 % Sat., Flags, B532 SD,...

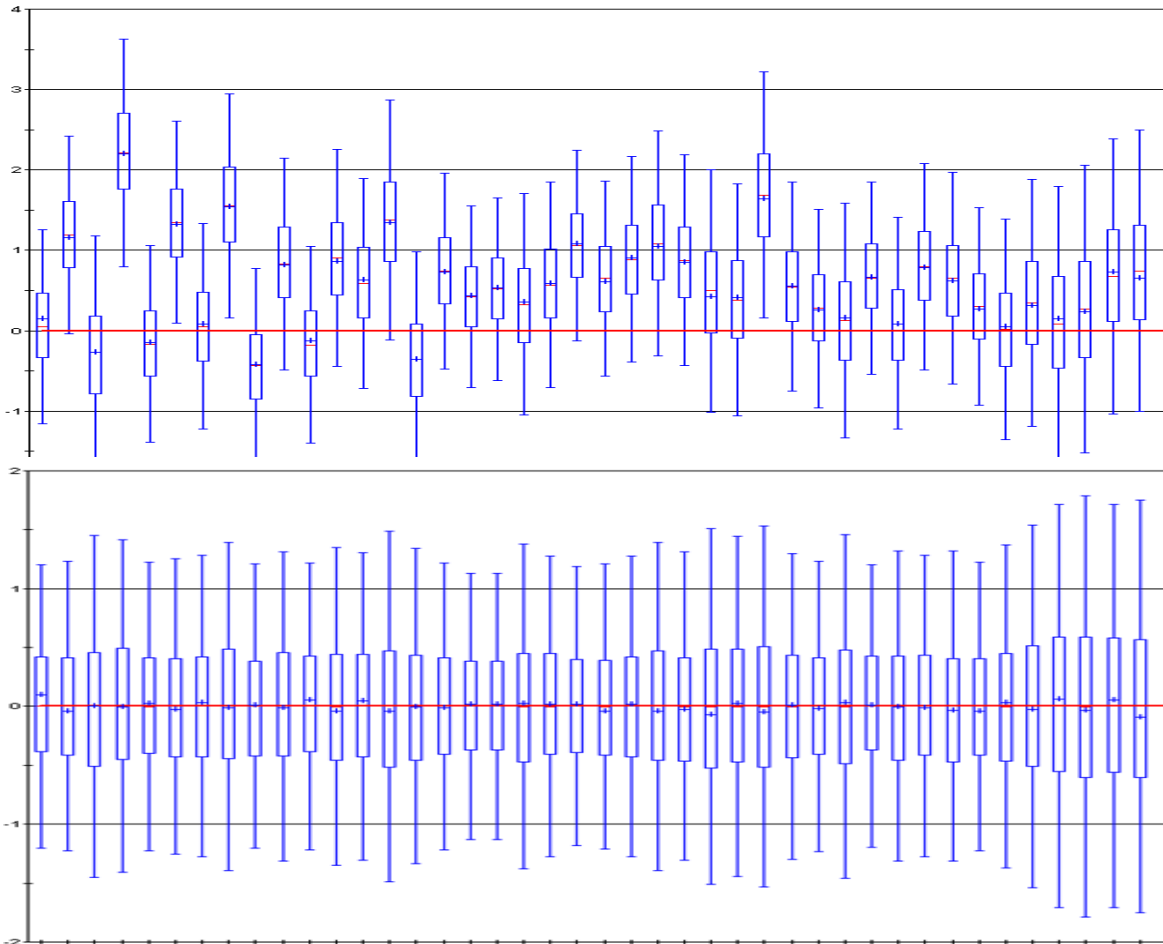


Normalization

- Removal of all sources of systematic non-biological variability and the reduction of the random errors.
- Basic assumption is that most of the genes are not changing their expression during the studied process



Box plot

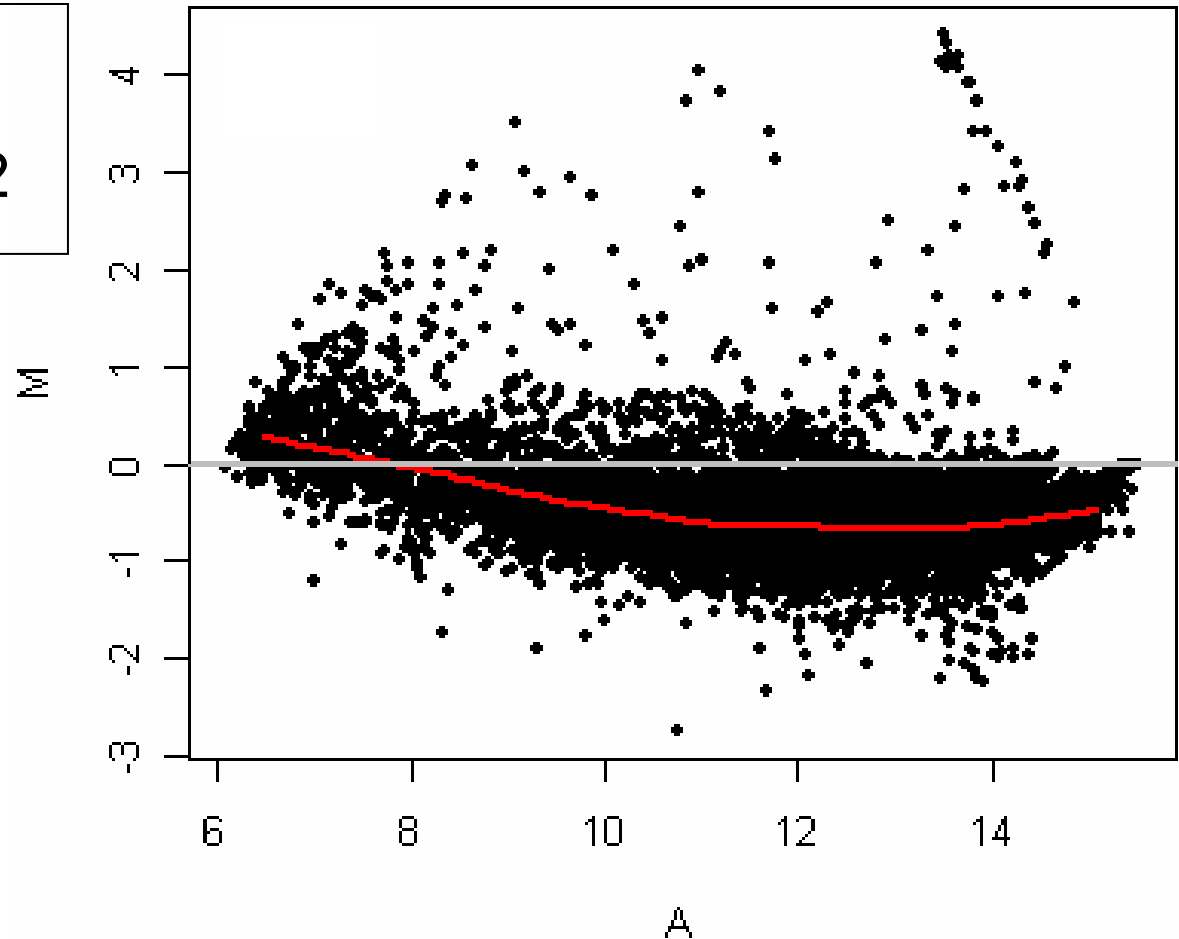




MA plot

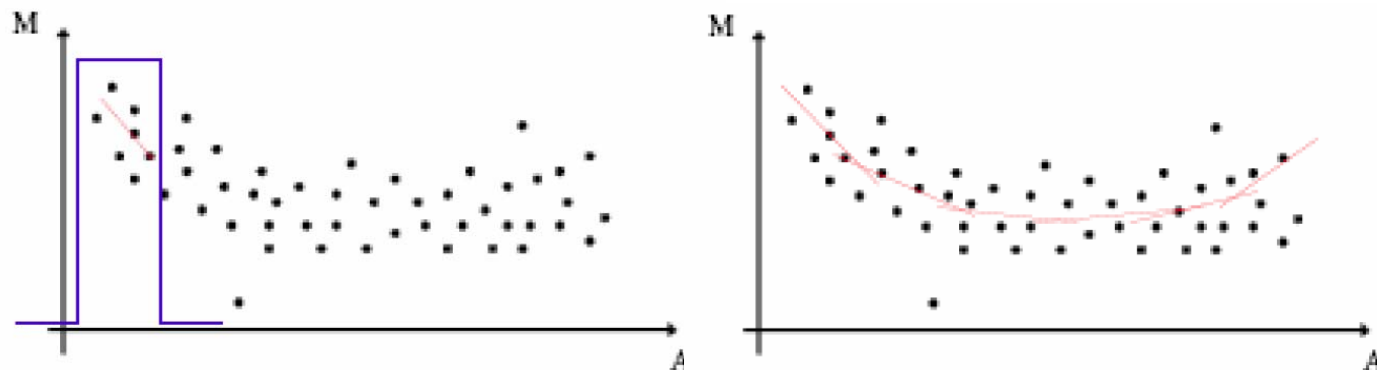
$$M = \log_2(R/G)$$

$$A = \log_2(R \cdot G)/2$$



Intensity dependent normalization

- Apply a locally weighted polynomial regression for a fixed subset of genes in the neighborhood of every gene i (LOWESS).



- Weight function:

$$w(x_i) = \begin{cases} (1 - d(x, x_i))^3 & : d(x, x_i) < 1 \\ 0 & : d(x, x_i) \geq 1 \end{cases}$$



Self normalization (dye-swap normalization)

$$M = \log_2 (R/G)$$
$$M' = \log_2 (R'/G')$$

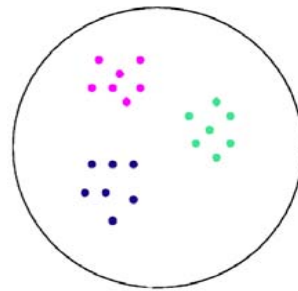
$$M_n \approx [\log_2 (R/G) - \log_2 (R'/G')] / 2$$

$$M_n = [\log_2 (RG'/GR')] / 2$$

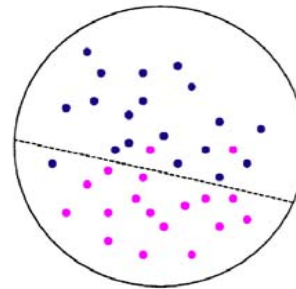


Clustering

- Unsupervised or supervised (classification)



Unsupervised



Supervised



Why cluster gene expression profiles

- Functionally related genes are often co-expressed
- Relationship between co-expression and co-regulation
- If a gene has unknown function, but clusters with genes of known function, this is a way to assign its general function ('guilt-by-association')



Methods for unsupervised clustering

- Hierarchical Clustering
- K-means
- ...



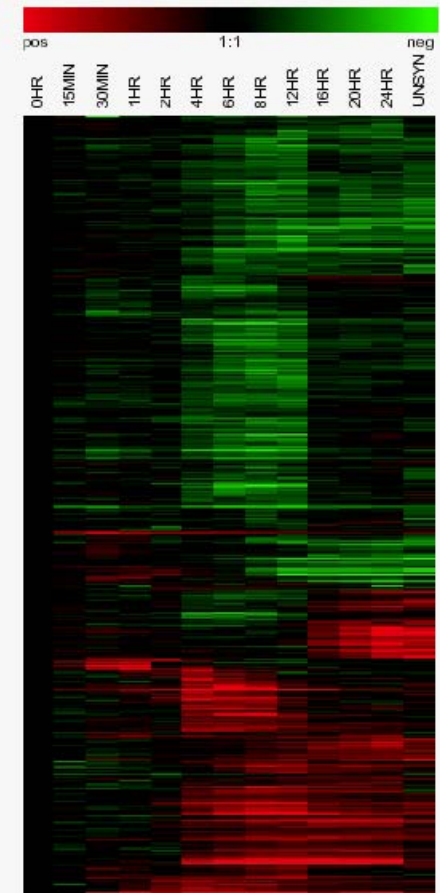
Data format

$(n \times m)$ Matrix of n Genes and m Experiments

$$x_{ij} = \log_2 \frac{C5_{ij}}{C3_{ij}}$$

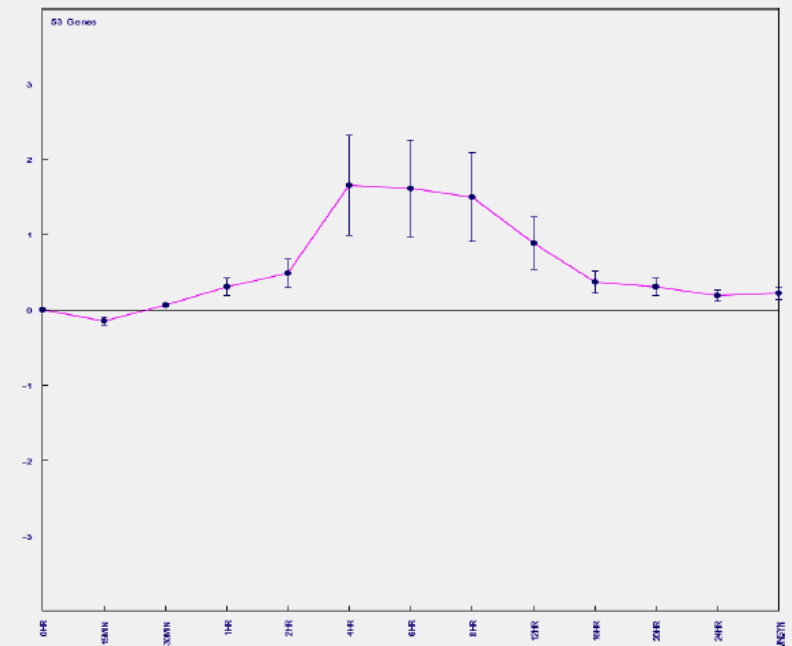
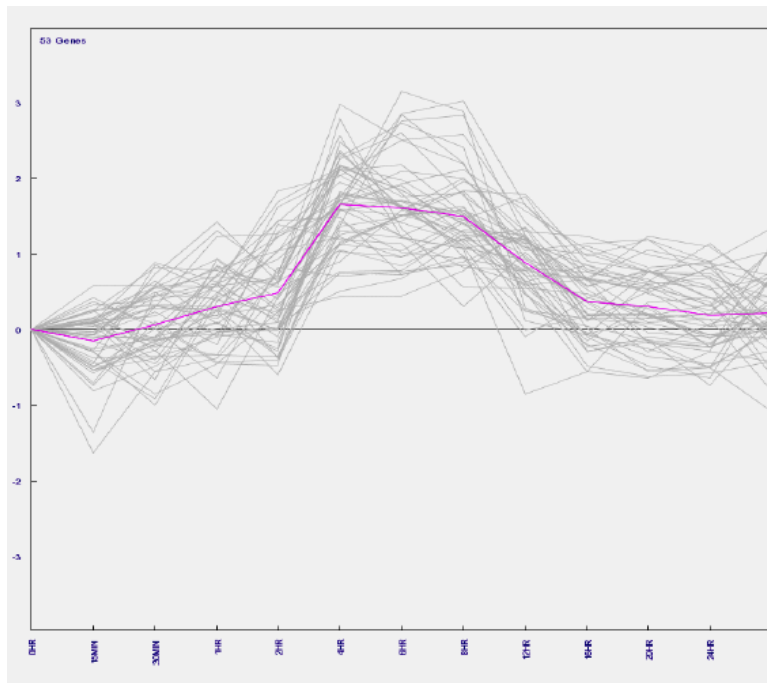
$C5_{ij}$...Cye-5 of gene i in microarray experiment j

$C3_{ij}$...Cye-3 of gene i in microarray experiment j





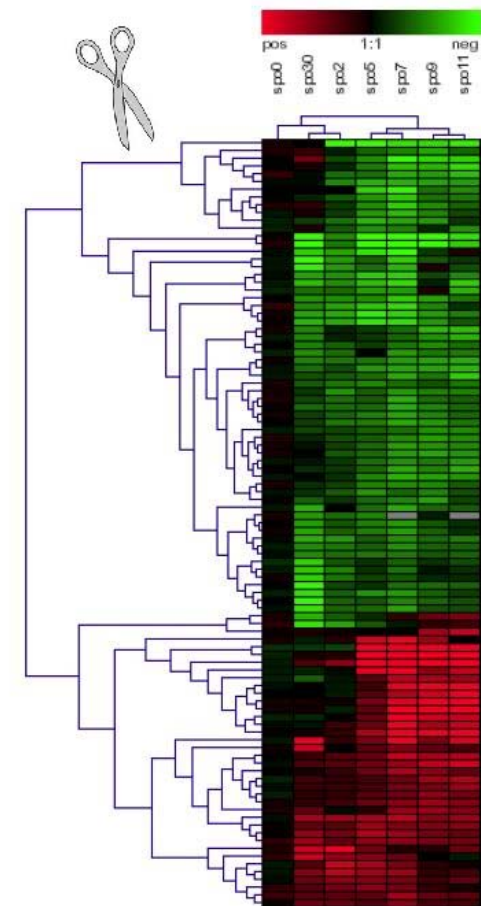
Graphical presentation





Hierarchical clustering

- Reorders the vectors regarding similarity
- Distances are encoded in dendrogram (tree)
- Unsupervised
- Very computational intensive
- Clusters of genes and experiments (bi-clustering)





Hierarchical clustering

Nodes = genes or groups of genes. Initially all nodes are rows of data matrix

- 1) Compute matrix of all distances (correlation coefficient)
- 2) Find two closest nodes.
- 3) Merge them by averaging measurements (weighted)
- 4) Compute distances from merged node to all others
- 5) Repeat until all nodes merged into a single node



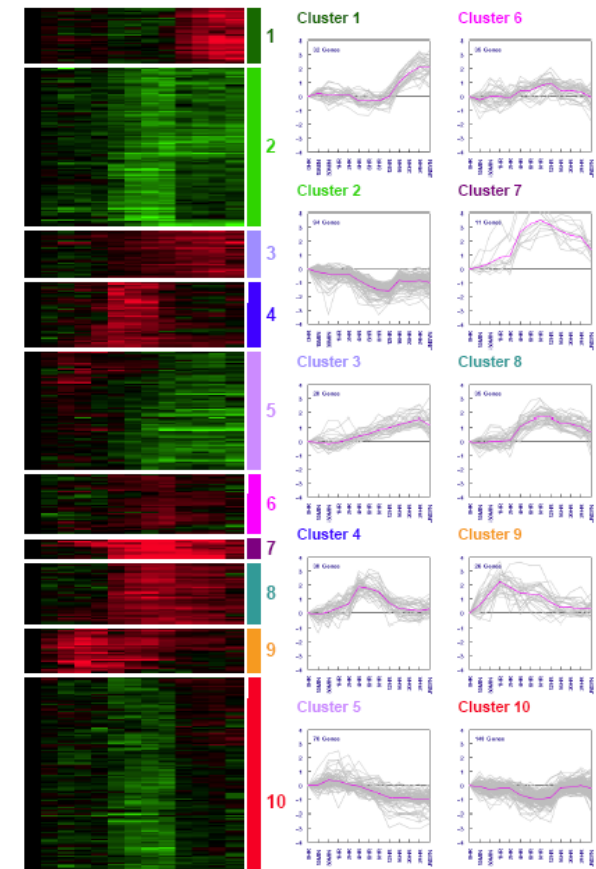
Linking within hierarchical clustering

- Single-linkage clustering
The distance between two clusters, i and j , is calculated as the minimum distance between a member of cluster i and a member of cluster j .
- Complete-linkage clustering
Here the maximum distance is used
- Average-linkage clustering
Calculated using average values (UPGMA)
- Weighted pair-group average
Like UPGMA but weighted according cluster size



K-means

- It partitions n genes into k clusters, where k has to be predetermined
- k-means clustering minimizes the variability within each cluster
- Tries to maximize the distance between clusters
- Moderate memory and time consumption





K-means

- 0) Choose number of clusters
- 1) Generate random points (“cluster centers”) in n dimensions (results are depending on these seeds)
- 2) Compute distance of each data point to each of the cluster centers.
- 3) Assign each data point to the closest cluster center.
- 4) Compute new cluster center position as average of points assigned.
- 5) Loop to (2), stop when cluster centers do not move very much.



Exercise

- 2 Parts:
 - Two colour microarrays
 - Microarray background correction, normalization, dye swaps, M values.
 - Clustering of interesting genes
 - K-means, hierarchical



Exercise Microarrays

- We will use Bioconductor – open source software for bioinformatics; R based
- We will use the following packages:
 - marray, limma, Biobase, OLIN, gplots
- Installing packages
 - From Bioconductor
 - `source("http://bioconductor.org/biocLite.R")`
 - `biocLite("marray");`
 - From R
 - `install.packages("gplots")`
- Load packages
 - `library("marray")`



Exercise Microarrays

- Useful functions
 - `getwd()/setwd()` – get/set working directory
 - `par(mfrow = c(2,2), mar = c(2,2,2,2))`
 - `read.marrayInfo()/read.Galfile()/read.GenePix()`
 - `backgroundCorrect2()/maNorm()`
 - `maPlot()/maBoxplot()/plot()`
 - `read.csv()/cor()/heatmap()/image()`
 - `hclust()/kmeans()`